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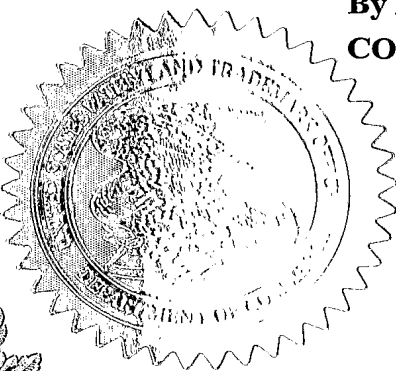
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**APPLICATION NUMBER: 60/559,452****FILING DATE: April 06, 2004**

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# PROVISIONAL APPLICATION FOR PATENT COVER SHEET

This is a request for filing a PROVISIONAL APPLICATION FOR PATENT under 37 CFR 1.53(c).

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PTO/SB/16 (01-04)

22856 U.S. PTO  
60/559452



INVENTOR(S)					
Given Name (first and middle [if any])		Family Name or Surname		Residence (City and either State or Foreign Country)	
Gerhard J.		Gries		Coquitlam, BC, Canada	
Additional inventors are being named on the _____ separately numbered sheets attached hereto					
TITLE OF THE INVENTION (500 characters max)					
Pheromone for manipulating codling moth larvae					
Direct all correspondence to: CORRESPONDENCE ADDRESS					
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OR					
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Country		Telephone		Fax	
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ENCLOSED APPLICATION PARTS (check all that apply) 4246					
<input type="checkbox"/> Specification Number of Pages _____		<input type="checkbox"/> CD(s), Number _____			
<input type="checkbox"/> Drawing(s) Number of Sheets _____		<input type="checkbox"/> Other (specify) _____			
<input type="checkbox"/> Application Data Sheet. See 37 CFR 1.76					
METHOD OF PAYMENT OF FILING FEES FOR THIS PROVISIONAL APPLICATION FOR PATENT					
<input checked="" type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27.				FILING FEE Amount (\$)	
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[Page 1 of 2]

Respectfully submitted,

SIGNATURE

TYPED or PRINTED NAME

TELEPHONE

*Gerhard Gries*  
Gerhard Gries  
604-944-4246

Date

REGISTRATION NO.

(if appropriate)

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Regine M.	Gries	Coquitlam, BC, Canada
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[Page 2 of 2]

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# FEE TRANSMITTAL for FY 2004

Effective 10/01/2003. Patent fees are subject to annual revision.

☒ Applicant claims small entity status. See 37 CFR 1.27

TOTAL AMOUNT OF PAYMENT

(\$)  
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## Complete if Known

Application Number

Filing Date

First Named Inventor

Zaid Jumean

Examiner Name

Art Unit

Attorney Docket No.

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1001 770	2001 385	Utility filing fee	
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1005 160	2005 80	Provisional filing fee	80.00
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### 2. EXTRA CLAIM FEES FOR UTILITY AND REISSUE

Total Claims	Extra Claims	Fee from below	Fee Paid
Independent Claims	-20** =	X	
Multiple Dependent	-3** =	X	

Large Entity Fee Code (\$)	Small Entity Fee Code (\$)	Fee Description
1202 18	2202 9	Claims in excess of 20
1201 86	2201 43	Independent claims in excess of 3
1203 290	2203 145	Multiple dependent claim, if not paid
1204 86	2204 43	** Reissue independent claims over original patent
1205 18	2205 9	** Reissue claims in excess of 20 and over original patent

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Large Entity Fee Code (\$)	Small Entity Fee Code (\$)	Fee Description	Fee Paid
1051 130	2051 65	Surcharge - late filing fee or oath	
1052 50	2052 25	Surcharge - late provisional filing fee or cover sheet	
1053 130	1053 130	Non-English specification	
1812 2,520	1812 2,520	For filing a request for <i>ex parte</i> reexamination	
1804 920*	1804 920*	Requesting publication of SIR prior to Examiner action	
1805 1,840*	1805 1,840*	Requesting publication of SIR after Examiner action	
1251 110	2251 55	Extension for reply within first month	
1252 420	2252 210	Extension for reply within second month	
1253 950	2253 475	Extension for reply within third month	
1254 1,480	2254 740	Extension for reply within fourth month	
1255 2,010	2255 1,005	Extension for reply within fifth month	
1401 330	2401 165	Notice of Appeal	
1402 330	2402 165	Filing a brief in support of an appeal	
1403 290	2403 145	Request for oral hearing	
1451 1,510	1451 1,510	Petition to institute a public use proceeding	
1452 110	2452 55	Petition to revive - unavoidable	
1453 1,330	2453 665	Petition to revive - unintentional	
1501 1,330	2501 665	Utility issue fee (or reissue)	
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1460 130	1460 130	Petitions to the Commissioner	
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1806 180	1806 180	Submission of Information Disclosure Stmt	
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1809 770	2809 385	Filing a submission after final rejection (37 CFR 1.129(a))	
1810 770	2810 385	For each additional invention to be examined (37 CFR 1.129(b))	
1801 770	2801 385	Request for Continued Examination (RCE)	
1802 900	1802 900	Request for expedited examination of a design application	

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Signature

Date

22 March 04

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March 25, 2004

In the United States Patent and Trademark Office

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Each of the undersigned understands:

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- C. Any desired foreign Convention application (including PCT applications) based upon this PPA must be filed within one year of the filing date of this PPA;
- D. This PPA must contain written description of the invention, and of the manner and process of making and using it, in such full, clear, concise, and exact term as to enable any person skilled in the art to which it pertains, or with which it is most nearly connected, to make and use the same, and shall set forth the best mode contemplated by the inventor of carrying out his invention, 35 U.S.C. §. 112, ¶ 1. otherwise this PPA will be worthless.
- E. Any RPA will be entitled to claim the benefit of the PPA only if such RPA names at least one inventor of this PPA and this PPA discloses such inventor's invention, as claimed in at least one claim of the RPA, in the matter provided in Item D above.

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**Title: Pheromone for manipulating codling moth larvae**

☒ Specification, Sheets: 20

☒ Drawings, Sheets: 12

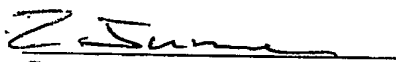
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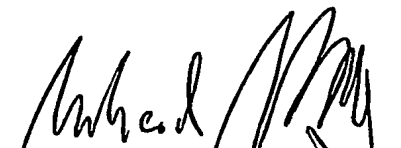
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March 25, 2004

Synthetic Aggregation Pheromone for Manipulating the Behaviour  
of Codling Moth, *Cydia pomonella*, Larvae

Zaid Jumean, Regine Gries, and Gerhard Gries



### Field of the Invention

This invention relates to a composition and procedure for manipulating the behaviour of codling moth larvae, *Cydia pomonella* (L.) (Lepidoptera: Olethreutidae). In particular, this invention relates to the use of specific pheromone components for manipulating the behaviour of *C. pomonella* larvae.

### Background of the Invention

Larvae of the codling moth, *Cydia pomonella* (L.) (Lepidoptera: Olethreutidae), feed on and cause damage to apple, pear, walnut and other fruit and nut crops (Pedigo 1999). In a typical apple orchard, if left untreated, *C. pomonella* larvae can infest up to 95% of the crop resulting in major economic loss (Pedigo 1999).

In temperate regions, larvae feed from June to August within (apple) fruits (Dolstad 1985). In August, larvae exit fruits and seek pupation sites, often on trunks of fruit-bearing trees. Spinning cocoons in which to pupate, larvae produce an aggregation pheromone that attracts or arrests other *C. pomonella* larvae (Duthie *et al.* 2003).

Components of this aggregation pheromone also attract the parasitic wasp *Mastrus ridibundus* (Hymenoptera: Ichneumonidae) which parasitize *C. pomonella* prepupae inside cocoons (Jumean *et al.*, unpublished). Synthetic aggregation pheromone in trapping devices would allow behavioural manipulation of *C. pomonella* larvae.

There are several patents listed in the United States Patent and Trademark Office database under the keyword *Cydia pomonella*. Two patents are concerned with the

synthesis of attractants for *C. pomonella*, as follows: U.S. Pat. No. 3,943,157 "Synthesis of codling moth attractant" reports the synthesis of codling moth sex pheromone, and U.S. Pat. No. 5,599,848 "Preparation, intermediates for the preparation and use of a mixture of dodecadienol isomers" reports a process for preparing and using the mixture of 8*E*,10*E*-dodecadienol, 8*E*,10*Z*-dodecadienol, 8*Z*,10*E*-dodecadienol, and 8*Z*,10*Z*-dodecadienol for interference of mating of *C. pomonella* adults. Three additional patents are concerned with methods of interfering with mating of *C. pomonella* adults, as follows: U.S. Pat. No. 6,395,775 "Combating pest insects" reports the use of the sex pheromone *E*8,*E*10-dodecadien-1-ol in combination with one or more behavioural antagonists or behavioural synergists for *C. pomonella* control. U.S. Pat. No. 4,734,281 "Method for concurrently emitting vapours of sex pheromones of different insects" reports the use of sex pheromone dispensers for controlling the population of two or more species of insect pests in the field, including *C. pomonella*. Finally, U.S. Pat. No. 6,528,049 "Bisexual attractants, aggregants, and arrestants for adult and larvae of codling moth and other species of lepidoptera" reports a method for monitoring and control of *C. pomonella* using attractants and arrestants from pears or apples. All of the behaviour-modifying compounds claimed for control of *C. pomonella* in the patents referred to above are very different from the attractive pheromone components produced by *C. pomonella* larvae, claimed in this application for attraction or arrestment of *C. pomonella* larvae that forage for suitable pupation sites.

### Summary of the Invention

We reveal pheromone components which attract or arrest male and female *C. pomonella* larvae. These pheromone components are derived from silk produced when either male or female larvae spin cocoons.

The essence of the invention is the preparation and implementation of these pheromone components for manipulating the behaviour of *C. pomonella* larvae. Pheromone components can be used in all possible combinations and ratios. Pheromone component compositions can be contained in slow release devices. Devices can be held in traps to retain male and female *C. pomonella* larvae. The invention can be used in combination with other tactics employed to control *C. pomonella* adults for protection of apple, pear, walnut, and other fruit and nut crops from *C. pomonella*.

The invention is directed to a composition of chemicals for manipulating the behaviour of *C. pomonella* larvae, said composition comprising pheromone components in all possible combinations and ratios selected from the group consisting of: 1) heptanal; 2) sulcatone; 3) myrcene; 4) octanal; 5) 3-carene; 6) (+)-limonene; 7) (*E*)-2-octenal; 8) nonanal; 9) (*E*)-2-nonenal; 10) decanal; and 11) geranylacetone.

The composition can be contained in, or released from, slow release devices. The composition can be contained in, or released from, a trap that captures attracted *C. pomonella* larvae.

The invention is also directed to an apparatus for attracting or arresting *C. pomonella* larvae, said apparatus containing a composition comprising pheromone components in all possible combinations and ratios selected from the group consisting of: 1) heptanal; 2) sulcatone; 3) myrcene; 4) octanal; 5) 3-carene; 6) (+)-limonene; 7) (*E*)-2-octenal; 8) nonanal; 9) (*E*)-2-nonenal; 10) decanal; and 11) geranylacetone.

The invention is also directed to a bait and apparatus for deployment in orchards susceptible to *C. pomonella*, said bait incorporating pheromone components in all possible combinations and ratios and an apparatus which is suitable for *C. pomonella* larvae to pupate in.

The invention also pertains to a method of manipulating the behaviour of *C. pomonella* larvae which comprises exposing the insects to one or more pheromone components according to the invention.

The invention also pertains to a method of diagnosing whether protection of an apple, pear, walnut or other fruit or nut crop is warranted, comprising exposing apple, pear, walnut, or other fruit or nut crops to a composition of two or more pheromone components according to the invention, and determining whether any *C. pomonella* larvae are attracted or arrested by the composition of pheromone components.

The invention includes a method of protecting apple, pear, walnut, or other fruit or nut crops from attack by *C. pomonella* larvae by deploying proximate to apple or pome fruit crops the composition of pheromone components according to the invention.

### Drawings

Drawings illustrate specific embodiments of the invention, but should not be construed as restricting the spirit or scope of the invention in any way:

FIG. 1 illustrates graphical data of responses of male or female *Cydia pomonella* larvae to pitfall devices baited with 1-day-old *C. pomonella* cocoons containing a male or female *C. pomonella* larva/prepupa.

FIG. 2 illustrates flame ionization detector (FID) and electroantennographic detector (EAD, female *Mastrus ridibundus* antenna) responses to ten cocoon-spinning *Cydia pomonella* larvae hour equivalents of cocoon volatile extracts.

FIG. 3 illustrates graphical data of responses of *Cydia pomonella* larvae to pitfall devices baited with natural or synthetic pheromone components from cocoon spinning *C. pomonella* larvae.

FIG. 4 illustrates graphical data of responses of *Cydia pomonella* larvae to pitfall devices baited with a synthetic 11-component pheromone blend at various doses.

FIG. 5 illustrates graphical data of responses of *Cydia pomonella* larvae to pitfall devices baited with synthetic pheromone blends lacking specific classes of pheromone components.

FIG. 6 illustrates graphical data of responses of *Cydia pomonella* larvae to pitfall devices baited with synthetic pheromone blends lacking individual pheromone components or specific groups of pheromone components.

FIG. 7 illustrates graphical data of responses of *Cydia pomonella* larvae to pitfall devices baited with various blends of synthetic pheromone components.

FIG. 8 illustrates graphical data of responses of *Cydia pomonella* larvae to pitfall devices baited with a synthetic 11-component pheromone blend, with components at natural ratios or increased amounts (x 10) of specific components.

FIG. 9 illustrates graphical data of responses of *Cydia pomonella* larvae in on-tree experiments to corrugated cardboard bands baited with live *C. pomonella* larvae/prepupae or synthetic pheromone blends.

FIG. 10 illustrates graphical data of responses of *Cydia pomonella* larvae in on-tree experiments to corrugated cardboard bands baited with synthetic pheromone blends at various doses.

FIG. 11 illustrates graphical data of responses of *Cydia pomonella* larvae in on-tree experiments to corrugated cardboard bands baited with synthetic pheromone blends, with specific components increased (x 10).

FIG. 12 illustrates graphical data of responses of *Cydia pomonella* larvae in on-tree experiments to corrugated cardboard bands baited with synthetic pheromone blends, with components at natural ratios or increased amounts (x 10) of specific components.

### Detailed Description of the Invention

Throughout the following description, specific details are set forth in order to provide a more thorough understanding of the invention. However, the invention may be practiced without these particulars. In other instances, well known elements have not been shown or described in detail to avoid unnecessarily obscuring the invention. Accordingly, the specification and drawings are to be regarded in an illustrative, rather than a restrictive, sense.

#### 1. Response of male and female *Cydia pomonella* larvae to cocoon-spinning male or female larvae

Response of 5<sup>th</sup> instar *C. pomonella* larvae seeking pupation sites was tested in 2-choice Petri dish olfactometers (Duthie *et al.*, 2003), with modified Eppendorf tubes in pitfall devices preventing physical contact of larvae with test stimuli. For each of at least 30 replicates per experiment, one male or female larva was released in the center of the olfactometer, and its pupation site recorded 18-24 hours later. Test stimuli consisted of a

control corrugated cardboard (CB) strip (2.5 x 2.5 cm) or a treatment CB strip carrying five 1-day-old cocoons each containing either a male or female *C. pomonella* larva/prepupa.

Male and female *C. pomonella* larvae preferred to spin cocoons in pitfall devices baited with CB carrying five 1-day-old cocoons, with no preference for cocoons containing either male or female larvae/prepupae (FIG. 1).

FIG. 1 illustrates graphical data of responses of male or female *Cydia pomonella* larvae to pitfall devices baited with control corrugated cardboard (CB) strips or treatment CB strips carrying five 1-day-old cocoons containing either male or female *C. pomonella* larvae/prepupae. Asterisks on bars indicate a significant response to a particular treatment; Chi-square goodness of fit test with Yates' correction for continuity; \*  $P < 0.025$ ; \*\*  $P < 0.001$ .

## 2. Acquisition, analyses, and bioassays of pheromone components produced by cocoon-spinning *Cydia pomonella* larvae

To capture airborne pheromone components from cocoon-spinning larvae, three-hundred 5<sup>th</sup> instar male and female larvae were placed in a cylindrical Pyrex glass chamber (15.5 x 20 cm). An empty chamber served as control. A water aspirator drew charcoal-filtered air at ~2 l/min through each chamber and through a glass column (140 x 1.3 mm OD) containing Porapak Q (50-80 mesh, Waters Associates, Inc., Milford, Massachusetts 01757). After 72 hours, the filters were desorbed with 3 ml of pentane and ether (95:5).



Extracts were concentrated under a nitrogen stream so that 1  $\mu$ L was equivalent to ca. 10 cocoon-spinning larvae hour equivalents (10 CSLHE = volatiles released from 10 cocoon-spinning *C. pomonella* larvae during 1 hour).

Aliquots of 20 CSLHE of Porapak Q-captured volatiles were subjected to analysis by coupled gas chromatographic-electroantennographic detection (GC-EAD) (Arn *et al.* 1975) using an antenna of *Mastrus ridibundus*, a specialist parasitic wasp of *C. pomonella* prepupae, as the electroantennographic detector. Using antennae from female *M. ridibundus*, instead of *C. pomonella* larvae, in these analyses was necessary because *C. pomonella* larvae have antennae too small for electrophysiological studies. It was also justified because host-seeking female *M. ridibundus* respond to the pheromone produced by cocoon-spinning *C. pomonella* larvae. In these GC-EAD analyses, 10 components (8 visible in FIG. 2) elicited responses from *M. ridibundus* antennae.

FIG. 2 illustrates flame ionization detector (FID) and electroantennographic detector (EAD: female *Mastrus ridibundus* antenna) responses to 10 CSLHE of Porapak Q extract. Chromatography: Hewlett Packard (HP) 5890A gas chromatograph equipped with a fused silica column (30 m x 0.32 mm ID) coated with DB-23 (J & W Scientific, Folsom, California 95630, USA); linear flow velocity of carrier gas: 35cm/sec; injector and FID detector temperature: 240°C; temperature program: 1 min at 50°C, 10° C/min to 220°C. Full scan electron impact (EI) and chemical ionization (CI) mass spectra of EAD active compounds were obtained by GC-mass spectrometry (MS) using a Varian Saturn II Ion Trap GC-MS and a HP 5985B GC-MS, respectively, each fitted with the DB-210

or DB-5 column. Candidate pheromone components were identified as follows: 1. heptanal (0.85); 2. sulcatone (0.81); 3. myrcene (0.84); 4. octanal (0.94); 5. 3-carene (0.95); 6. (+)-limonene (13.00); 7. (*E*)-2-octenal (0.41); 8. nonanal (4.10); 9. (*E*)-2-nonenal (1.00); 10. decanal (1.40); 11. geranylacetone (0.50). Note: 1) 10 CSLHE = volatiles released from 10 cocoon-spinning larvae during 1 hour; 2) number in brackets refers to nanogram quantities present in 10 CSLHE; 3) (+)-limonene was not antennally active but was included in bioassay experiments as the most abundant component in extracts.

### 3. Response of *Cydia pomonella* larvae to blends of natural or synthetic candidate pheromone components in olfactometer experiments

In olfactometer experiments (following the general protocol as described above), *C. pomonella* larvae preferred Porapak Q volatile extract of cocoon-spinning *C. pomonella* larvae, and a synthetic blend (SB) of 11 candidate pheromone components, over a pentane control stimulus (FIG. 3).

FIG. 3 illustrates graphical data of responses of *Cydia pomonella* larvae to pitfall devices baited with Porapak Q volatile extract of cocoon-spinning larvae (180 CSLHE) or a synthetic blend (SB) of 11 candidate pheromone components. Asterisks on bars indicate a significant response to a particular treatment; Chi-square goodness of fit test with Yates' correction for continuity; \*\*\* $P < 0.001$ . Note: 1) 180 CSLHE = volatiles released from 180 cocoon-spinning *C. pomonella* larvae during 1 hour; 2) components in SB

consisted of: 1. heptanal; 2. sulcatone; 3. myrcene; 4. octanal; 5. 3-carene; 6. (+)-limonene; 7. (*E*)-2-octenal; 8. nonanal; 9. (*E*)-2-nonenal; 10. decanal; 11. geranylacetone.

Similar attractiveness of Porapak Q extract containing natural cocoon volatiles and the synthetic 11-component blend (SB) strongly suggested that all essential pheromone components were present in SB.

Of the 4 doses of SB (1, 10, 100, 1,000 CSLHE) bioassayed, 100 SB elicited the strongest response by *C. pomonella* larvae (FIG. 4).

FIG. 4 illustrates graphical data of responses of *Cydia pomonella* larvae in pitfall olfactometer experiments to 4 doses of a synthetic blend (SB) containing 11 candidate pheromone components. Bars with an asterisk indicate a significant preference for a particular treatment; Chi-square goodness of fit test with Yates' correction for continuity; \*\*  $P < 0.005$ . Note: components in SB consisted of: 1. heptanal; 2. sulcatone; 3. myrcene; 4. octanal; 5. 3-carene; 6. (+)-limonene; 7. (*E*)-2-octenal; 8. nonanal; 9. (*E*)-2-nonenal; 10. decanal; 11. geranylacetone.

To determine the critically important components in SB, individual or groups of pheromone components were deleted and such reduced blends bioassayed in olfactometer experiments (following the protocol described above).

SB lacking monoterpenes [(+)-limonene, myrcene, 3-carene] or ketones (sulcatone, geranylacetone ) still elicited responses from *C. pomonella* larvae, whereas SB lacking aldehydes [heptanal, octanal, nonanal, decanal, (*E*)-2-octenal, (*E*)-2-nonenal] was completely unattractive (FIG. 5).

FIG. 5 illustrates graphical data of responses of *Cydia pomonella* larvae to pitfall devices baited with synthetic blends (SB) lacking either ketones [sulcatone, geranylacetone], monoterpenes [(+)-limonene, myrcene, 3-carene], or aldehydes [heptanal, octanal, nonanal, decanal, (*E*)-2-octenal, (*E*)-2-nonenal]. Asterisks on bars indicate a significant preference for a particular treatment; Chi-square goodness of fit test with Yates' correction for continuity; \*  $P < 0.05$ ; \*\*  $P < 0.01$ .

SBs lacking saturated aldehydes [heptanal, octanal, nonanal, decanal] still elicited significant responses from *C. pomonella* larvae, whereas SBs lacking either unsaturated aldehydes [(*E*)-2-octenal, (*E*)-2-nonenal], sulcatone, or geranylacetone were as ineffective as pentane controls in eliciting response from *C. pomonella* larvae (FIG. 6).

FIG. 6 illustrates graphical data of responses of *Cydia pomonella* larvae to synthetic blends (SB) lacking either saturated aldehydes [heptanal, octanal, nonanal, decanal], unsaturated aldehydes [(*E*)-2-octenal, (*E*)-2-nonenal], or individual components. The asterisk on bars indicates a significant preference for a particular treatment; Chi-square goodness of fit test with Yates' correction for continuity; \*  $P < 0.01$ .

A rudimentary synthetic blend (RSB), comprising (*E*)-2-octenal, (*E*)-2-nonenal, sulcatone and geranylacetone, did not elicit a behavioural response by *C. pomonella* larvae.

However, RSB in combination with either the monoterpene 3-carene, saturated aldehydes (octanal, nonanal, decanal), or 3-carene plus said aldehydes, induced significant attraction/arrestment by *C. pomonella* larvae (FIG. 7).

FIG. 7 illustrates graphical data of responses of *Cydia pomonella* larvae to a 4-component rudimentary synthetic blend (RSB) [(*E*)-2-octenal, (*E*)-2-nonenal, sulcatone, geranylacetone] alone or in combination with either 3-carene, saturated aldehydes [octanal, nonanal, decanal], or both. Asterisks on bars indicate a significant preference for a particular treatment; Chi-square goodness of fit test with Yates' correction for continuity; \*  $P < 0.01$ ; \*\*  $P < 0.005$ .

The 11-component synthetic blend (SB), when tested at the low dose of 10 cocoon-spinning larval hour equivalents and at natural compound ratios (Fig. 2), did not induce behavioural responses by *C. pomonella* larvae. However, a 10-fold increase of both (*E*)-2-octenal and (*E*)-2-nonenal, but not of either component singly, in SB resulted in significant attraction/arrestment of *C. pomonella* larvae (FIG. 8).

FIG. 8 illustrates graphical data of responses by *Cydia pomonella* larvae to a synthetic blend (SB) of 11 components at natural ratios (Fig. 2), or at a 10-fold increase of either or both (*E*)-2-octenal and (*E*)-2-nonenal. The asterisk indicates a significant preference for a particular treatment; Chi-square goodness of fit test with Yates' correction for

continuity; \*  $P < 0.05$ . Note: SB was tested at 10 cocoon-spinning larval hour equivalents.

#### 4. On-tree testing of natural or synthetic larval aggregation pheromone

In on-tree experiments, maple (*Acer* spp.) trees (10-16 cm diameter at a height of 45 cm) were banded with corrugated cardboard strips (5 cm wide) 45 cm above ground. Strips were divided into two halves, with test stimuli applied to the waxed, center portion of each half. For each replicate in all experiments, twenty 5<sup>th</sup> instar *C. pomonella* larvae were released from a thin, circular platform affixed to the base of the tree's main branch crotch (~1.50 m above ground). Experiments were initiated after dusk, and numbers of *C. pomonella* larvae cocooning in treatment or control halves of corrugated cardboard strips recorded 10-12 hours later.

#### EXAMPLE # 1

In experiment 31, significantly more *C. pomonella* larvae cocooned in treatment halves of corrugated cardboard strips, bearing twenty-five 1-day-old *C. pomonella* cocoons with female larvae/prepupae inside, than in unbaited control halves. Similarly, in experiment 32 significantly more *C. pomonella* larvae cocooned in halves of corrugated cardboard strips baited with a synthetic blend (SB) of pheromone components than in halves treated with a solvent control (FIG. 9).

FIG. 9 illustrates graphical data of responses by *Cydia pomonella* larvae in on-tree experiments 31 (12 replicates) and 32 (18 replicates) to corrugated cardboard (CB) strips.

Treatment halves of CB strips carried twenty-five 1-day-old cocoons containing female *C. pomonella* larvae/prepupae (Exp. 31), or were baited with a synthetic blend (SB) of 11 components at 1,000 cocoon-spinning larval hour equivalents (Exp. 32). Control halves were bare (Exp. 31), or were impregnated with the equivalent amounts of solvent (Exp. 32). Asterisks indicate a significant preference for a particular stimulus; Wilcoxon paired-sample test; \*  $P < 0.01$ ; \*\*  $P < 0.005$ .

### EXAMPLE # 2

In on-tree experiments 33-35, the synthetic blend (SB) at 1,000 cocoon-spinning larvae hour equivalents (CSLHE), but not at 100 or 10,000 CSLHE, attracted/arrested significantly more *C. pomonella* larvae than did a solvent control stimulus (FIG. 10)

FIG. 10 illustrates graphical data of responses by *Cydia pomonella* larvae in on-tree experiments 33 (18 replicates), 34 (18 replicates) and 36 (12 replicates) to corrugated cardboard (CB) strips impregnated with a synthetic blend (SB) of 11 components at 100, 1,000, or 10,000 cocoon spinning larvae hour equivalents, or a solvent control. The asterisk indicates a significant preference for a particular stimulus; Wilcoxon paired-sample test; \*  $P < 0.01$ .

### EXAMPLE # 3

In on-tree experiment 36, significantly more *C. pomonella* larvae cocooned in halves of corrugated cardboard (CB) strips impregnated with a synthetic blend (SB) of 11 components, tested at 100 cocoon-spinning larval hour equivalents with increased

amounts (x 10) of (*E*)-2-octenal and (*E*)-2-nonenal, than on halves impregnated with a solvent control (FIG. 11).

FIG. 11 illustrates graphical data of responses by *Cydia pomonella* larvae in on-tree experiment 36 (12 replicates) to corrugated cardboard (CB) strips impregnated with a synthetic blend (SB) of pheromone components or a solvent control. The asterisk indicates a significant preference for a particular stimulus; Wilcoxon paired-sample test; \*  $P < 0.02$ . Note: SB was tested at 100 cocoon-spinning larval hour equivalents with increased amounts (x 10) of (*E*)-2-octenal and (*E*)-2-nonenal.

#### EXAMPLE # 4

In concurrently run on-tree experiments 37 and 38, significantly more *C. pomonella* larvae cocooned in halves of corrugated cardboard strips impregnated with a synthetic blend (SB) of 11 components at doses of 1,000 cocoon-spinning larval hour equivalents (CSLHE), or a modified synthetic blend at 100 CSLHE, than in halves impregnated with the equivalent amount of a solvent (pentane) control (FIG. 12).

FIG. 12 illustrates graphical data of responses by *Cydia pomonella* larvae in concurrently run on-tree experiments 37 (24 replicates) and 38 (24 replicates) to halves of corrugated cardboard (CB) strips impregnated with synthetic blends (SB) of 11 components. An asterisk indicates a significant preference for a particular stimulus; Wilcoxon paired-sample test; \*  $P < 0.005$ . Note: In experiment 38, the SB at 100 cocoon-spinning larvae hour equivalents contained increased amounts (x 10) of (*E*)-2-octenal and (*E*)-2-nonenal.



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## WHAT IS CLAIMED IS:

1. A composition of chemicals for manipulating the behaviour of *Cydia pomonella* larvae, said composition comprising two or more chemicals in all possible combinations and ratios selected from the group consisting of: 1) heptanal; 2) sulcatone; 3) myrcene; 4) octanal; 5) 3-carene; 6) (+)-limonene; 7) (*E*)-2-octenal; 8) nonanal; 9) (*E*)-2-nonenal; 10) decanal; 11) geranylacetone.
2. A composition as claimed in claim 1 wherein the composition is contained in, or released from, slow release devices.
3. A composition as claimed in claim 1 wherein the composition is contained in, and released from, an apparatus that provides suitable pupation sites for attracted *Cydia pomonella* larvae.
4. An apparatus for attracting *Cydia pomonella* larvae, said apparatus containing a composition comprising two or more chemicals in all possible combinations and ratios selected from the group consisting of: 1) heptanal; 2) sulcatone; 3) myrcene; 4) octanal; 5) 3-carene; 6) (+)-limonene; 7) (*E*)-2-octenal; 8) nonanal; 9) (*E*)-2-nonenal; 10) decanal; 11) geranylacetone.
5. An apparatus as claimed in claim 4 wherein the apparatus contains an insect-killing agent.
6. A bait and apparatus for deployment in an area containing fruit or nut crops, said bait incorporating a composition of chemicals as claimed in claim 1, and said apparatus providing suitable pupation sites for *Cydia pomonella* larvae.

7. A method of manipulating the behaviour of *Cydia pomonella* larvae which comprises exposing larvae to two or more chemicals as claimed in claim 1.

8. A method of diagnosing whether protection of apple, pear, walnut, or other fruit or nut crops is warranted, comprising exposing the fruit or nut crop to a composition as claimed in claim 1, and determining whether *Cydia pomonella* larvae are attracted by the composition.

9. A method of protecting apple, pear, walnut, or other fruit or nut crops from attack by *Cydia pomonella* larvae by deploying proximate to fruit or nut crops a composition as claimed in claim 1.

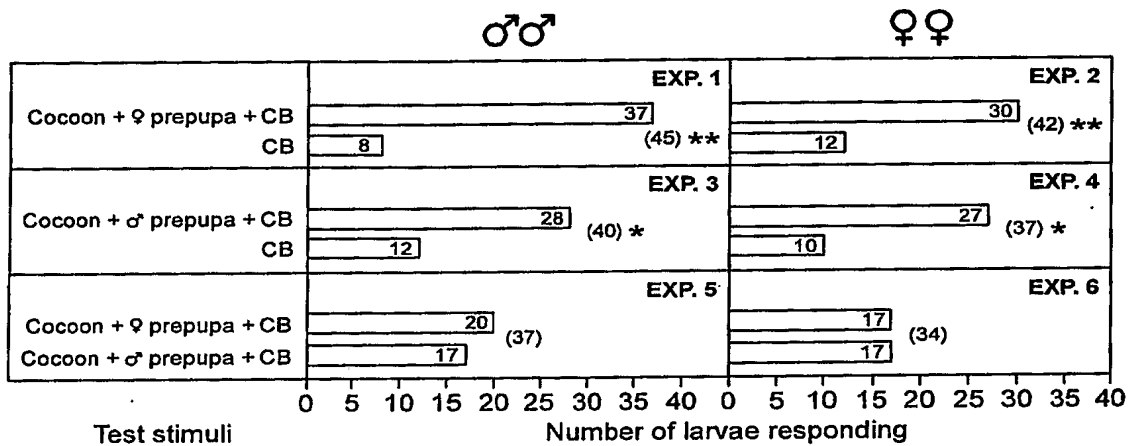


FIG. 1

Pomonella fig 1

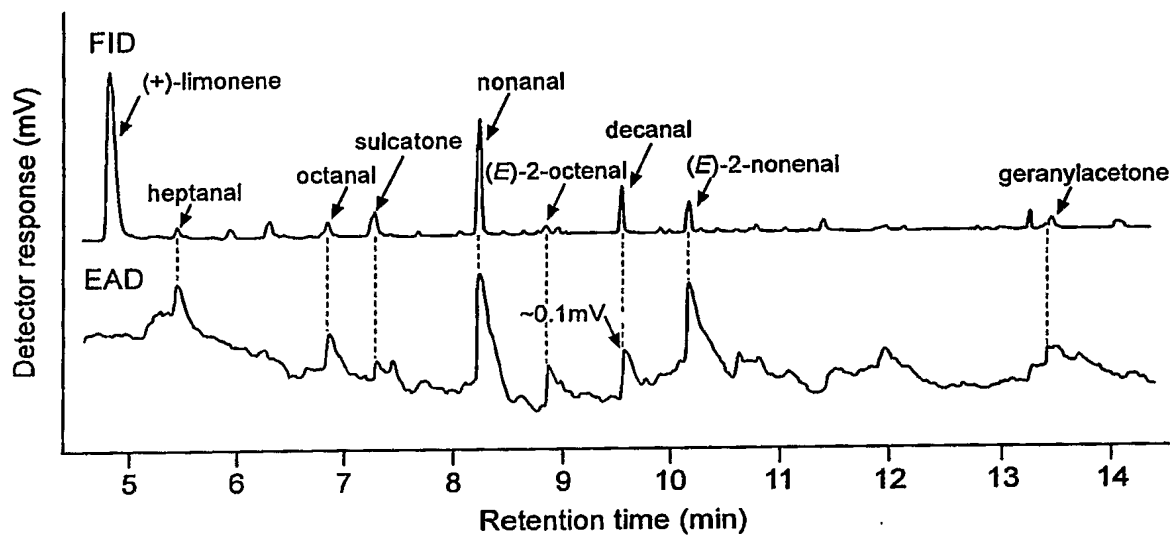


FIG. 2

Pomonella-ridi-fig 2

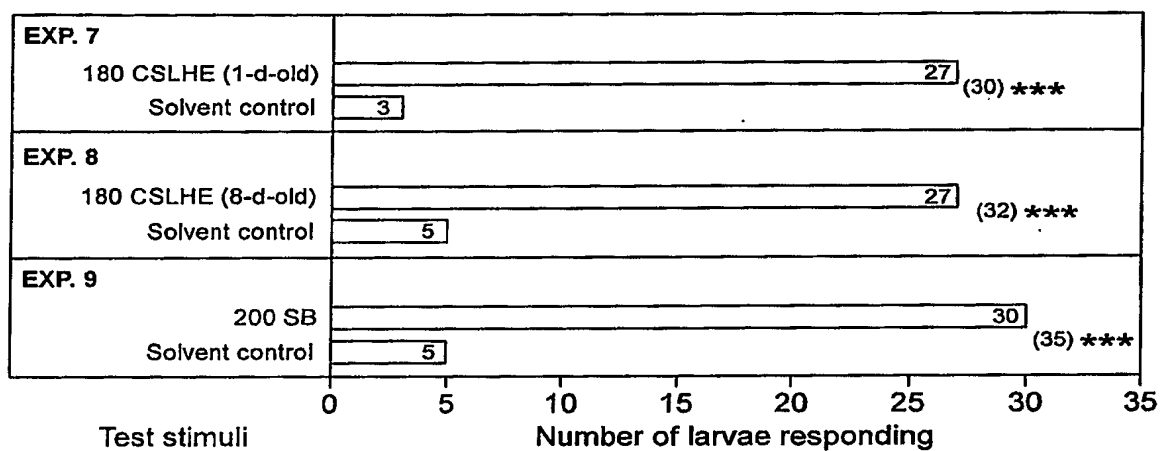
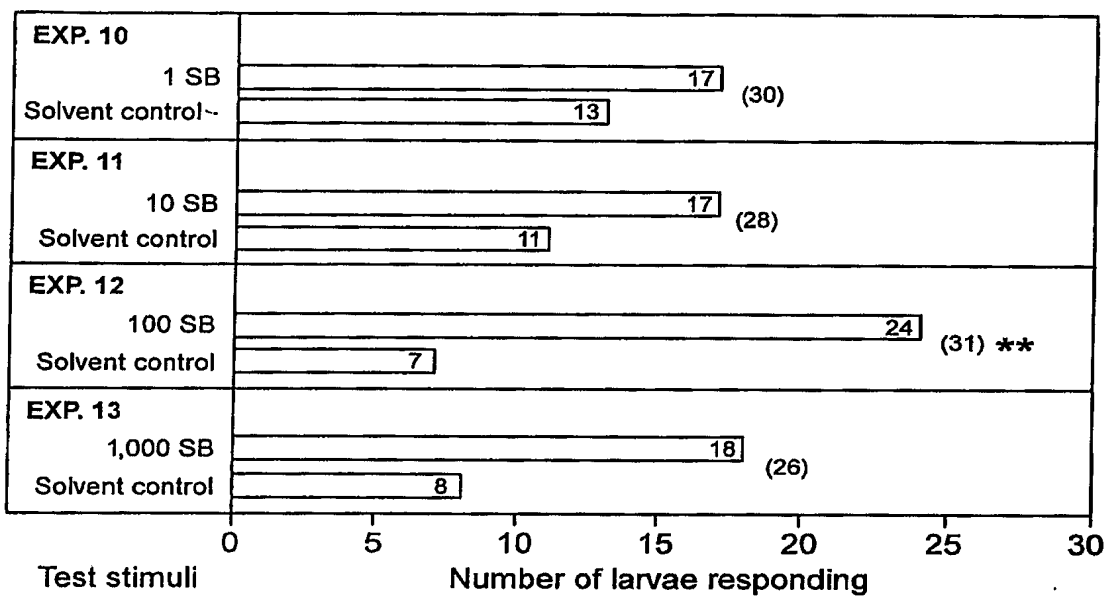


FIG. 3

Pomonella fig 3



**FIG. 4**

Pomonella fig 4

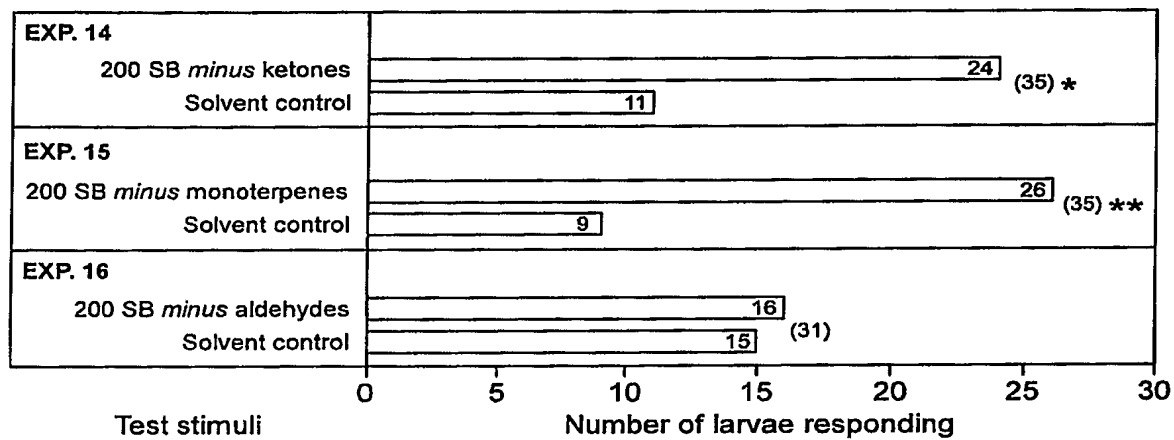


FIG. 5

Pomonella fig 5



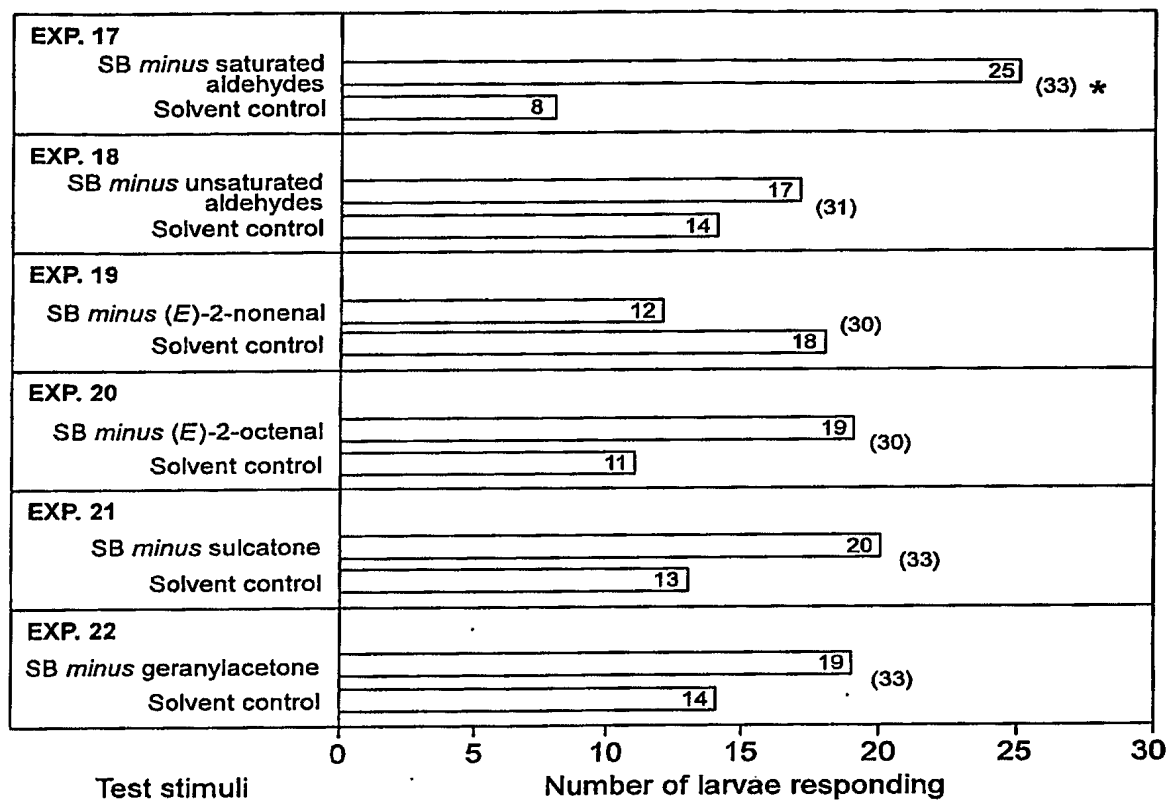


FIG. 6

Pomonella fig 6

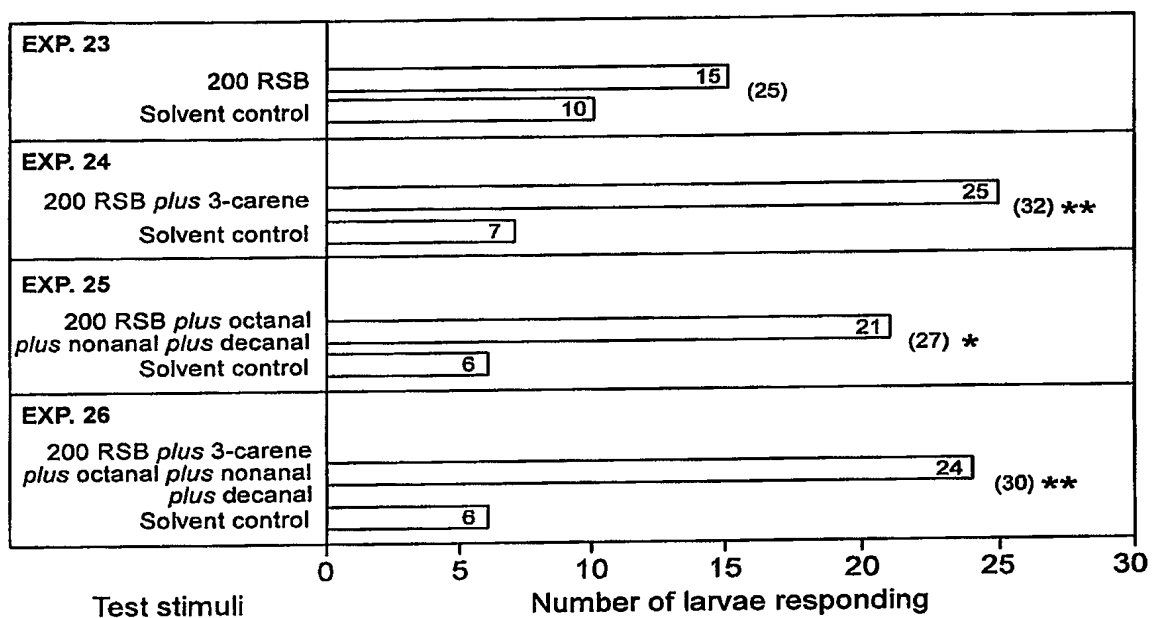


FIG. 7

Pomonella fig 7

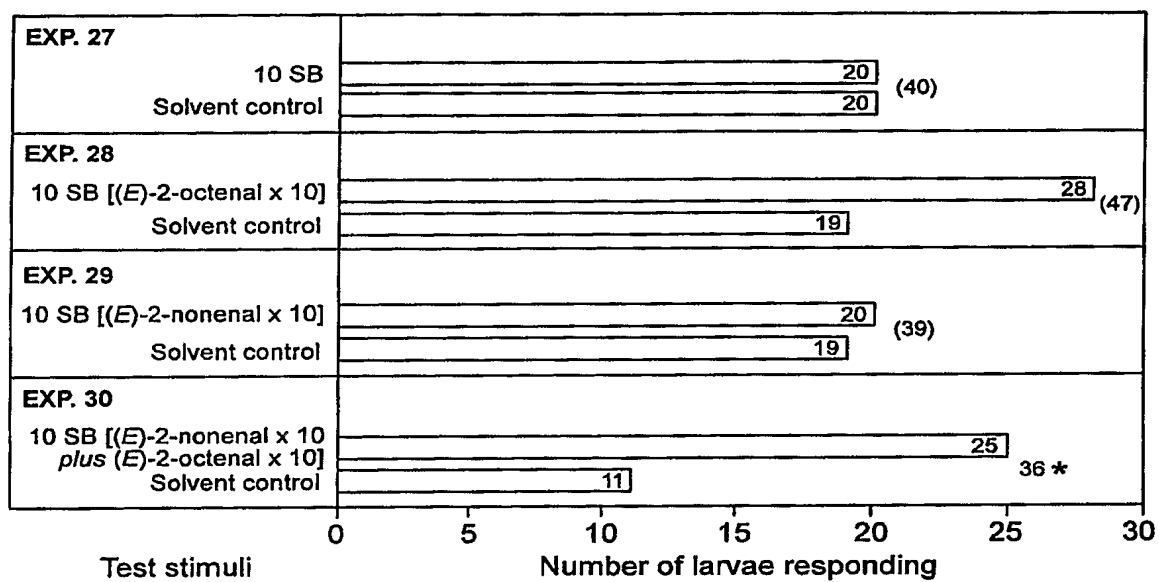


FIG. 8

Pomonella fig 8

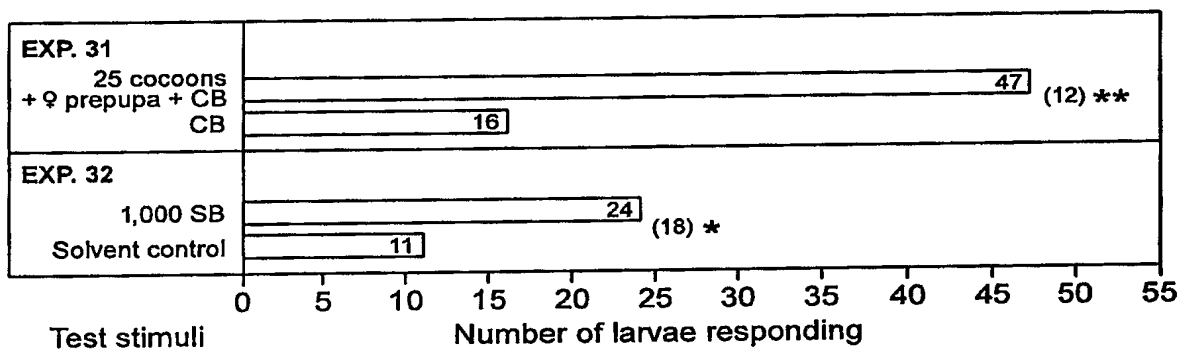


FIG. 9

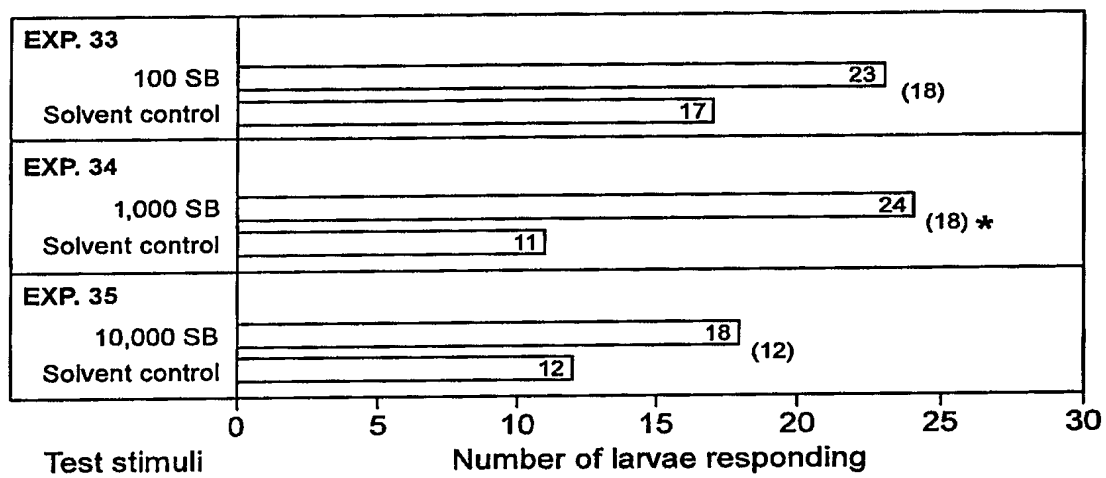


FIG. 10

Pomonella fig 10

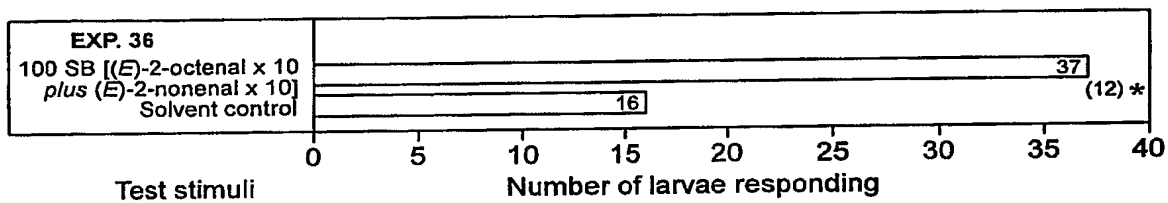


FIG. 11

Pomonella fig 11

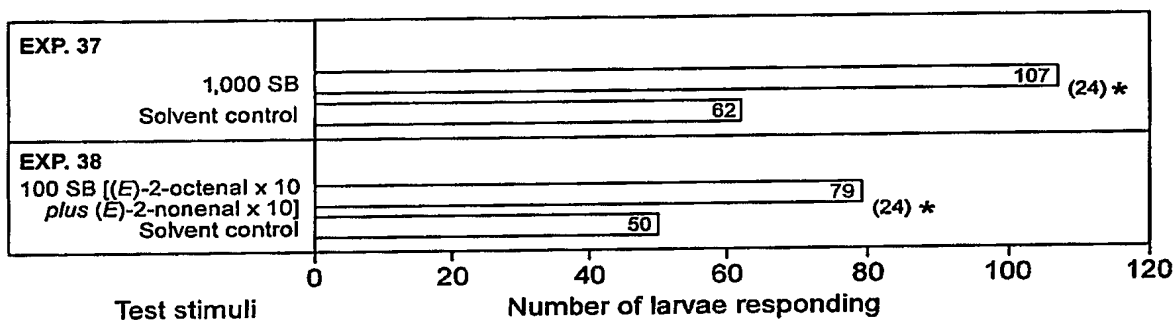


FIG. 12

Pomonella fig 12